

Cheminformatics of Drug-like Small Molecules

-
BioC2013

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Cheminformatics Basics

Structure Formats

Similarity Searching

Physicochemical Properties

Clustering

Hands-on Section

Compound Import/Export

Object Classes

Compound Structure Depictions

Compound Properties

Compound Similarity Searching

Compound Clustering

Outline

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Computations on Small Molecule Structures

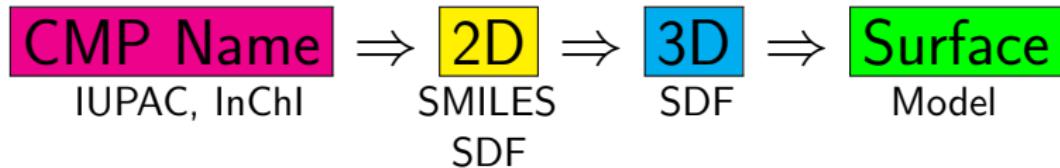
Requirements

- Computer readable representations of chemical structures

Challenges

- Compounds
 - Several connection types, many branch points and/or ring closures
- DNA/protein sequences
 - Linear strings, one connection type, usually no branch points or ring closures

Utility of Structure Formats



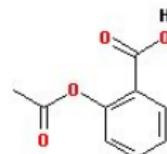
- Nomenclature to uniquely represent chemicals
- Computer representation and manipulation
- Format interconversions
- Representation of stereochemistry and 3D formats

Most Commonly Used Structure Formats

- Chemical nomenclature

- Trivial names: aspirin, acetylsalicylic acid
- IUPAC: 2-acetoxybenzoic acid
- InChI: 1.12Beta/C9H8O4/c1-6(10)13-8-5-3-2-4-7(8)9(11)12/h1H3,2-5H,(H,11,12)

Aspirin



- Line notations

- SMILES: CC(=O)Oc1ccccc1C(=O)O
- Other: WLN, ROSDAL, SLN, etc.

- Connection tables hold 3D & annotation information

- SDF (structure definition file)
- MDL Molfile
- Other: PDB, CML, etc.

Connection Table Formats: SDF and Mol

Molfile: header block and connection table (a, b)

SDfile: extension of Molfile (a, b, c)

(a) Header block

- (a1) CMP name or blank line
- (a2) software, date, 2/3D, ...
- (a3) blank line

(b) Connection table (CT)

- (b1) counts line: n atoms, n bonds, chiral, ...
- (b2) atom block: x,y,z coordinates, atoms, mass diff., charge, ...
2D representation when z coordinates all zero
- (b3) bond block: atom 1, atom 2, bond type, stereo specs, ...
- (b4) CT delimiter

(c) Annotation data

- (c1) <data header>
- (c2) data
- (c3) blank line
- (c4) continues like c1-3
- (c5) SDF delimiter (\$\$\$\$)

Example: SDF Format

```
a1      NSC85228 ethanol 1
a2      APtclserve02230600142D 0 0.00000 0.00000NCl NS
a3
b1      9 8 0 0 0 0 0 0 0 0999 V2000
b2      2.8660 -0.250 0.0000 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
b2      3.7321 0.2500 0.0000 C 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
b2      4.5981 -0.250 0.0000 C 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
b2      2.3291 0.0600 0.0000 H 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
b2      4.1306 0.7249 0.0000 H 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
b2      3.3335 0.7249 0.0000 H 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
b2      4.2881 -0.786 0.0000 H 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
b2      5.1350 -0.560 0.0000 H 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
b2      4.9081 0.2869 0.0000 H 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
b3      1 2 1 0 0 0 0
b3      2 3 1 0 0 0 0
b3      1 4 1 0 0 0 0
b3      2 5 1 0 0 0 0
b3      2 6 1 0 0 0 0
b3      3 7 1 0 0 0 0
b3      3 8 1 0 0 0 0
b3      3 9 1 0 0 0 0
b4      M END
c1      >< NSC >
c2      85228
c4      >< CAS >
c4      64-17-5
c4      >< SMILES >
c4      CCO
c5      $$$$
```

SMILES: Simplified Molecular Input Line Entry System

- Tutorial: <http://www.daylight.com/smiles/smiles-intro.html>
- Online rendering: <http://www.daylight.com/daycgi/depict>
- Non-canonical SMILES for manual entry
- Canonical SMILES needs to be computer generated
- Canonicalization: single ('correct') representation of several possibilities
 - OCC - ethanol
 - CCO - ethanol
- Canonical format important for databases

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Small Molecule Similarity Concepts

How to define similarities between compounds?

- Identical structure search
- Substructure and superstructure searches
- 2D fragment similarity searching
- 3D similarity searches (e.g. pharmacophore searching)
- Graph-based approaches (e.g. maximum common substructure: MCS)
- Many additional methods

2D Fragment Similarity Search Methods

Involve two major steps

- Encode structural descriptors from compounds
 - e.g. structural keys, fingerprints, atom pairs
- Similarity measure for encoded descriptors
 - e.g. Tanimoto coefficient, Euclidean

Structural Keys

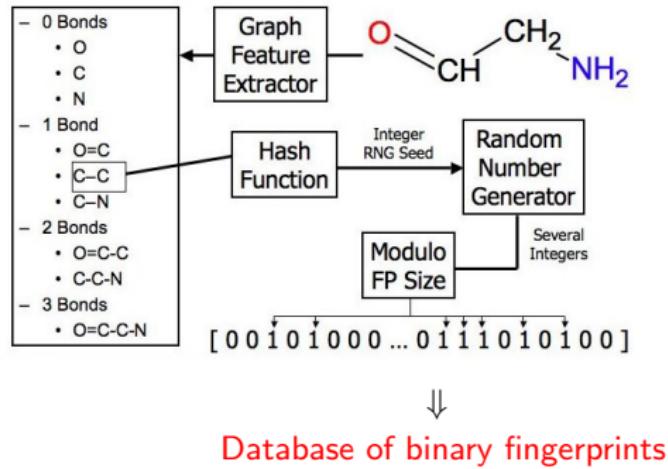
- Structural descriptors are based on lookup library of known "functional" substructures.
- Pre-compute presence of relevant substructures up front and encode them in bit-vector.
- Example of structural keys:
 - Presence of atoms (C, N, O, S, Cl, Br, etc.)
 - Ring systems
 - Aromatic, Phenol, Alcohol, Amine, Acid, Ester, ...
- Disadvantages:
 - Lookup library tends to be incomplete.
 - Sparsely populated vectors.

Fingerprints

- Fingerprints are generated directly from the molecule itself and not from a reference set of substructures.
- The algorithm examines each molecule and generates the following patterns:
 - One for each atom.
 - One representing each atom and its nearest neighbors (plus the bonds that join them).
 - One representing each group of atoms and bonds connected by paths up to 2, 3, 4, ... bonds long.
 - For example, the molecule OC=CN would generate the following patterns:
 - 0-bond paths: C, O, N
 - 1-bond paths: OC, C=C, CN
 - 2-bond paths: OC=C, C=CN
 - 3-bond paths: OC=CN,

Fingerprints

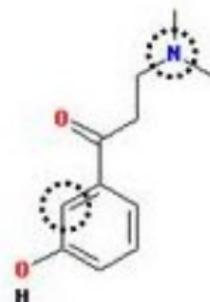
- Patterns are often encoded into fixed length (binary) vectors for fast similarity searching.
- Abstract, hard to traceback meaning of individual bits.



Atom Pair and Atom Sequence Similarity Searching

- Like fingerprints atom pairs are generated directly from the molecule itself and not from a reference set of substructures (Chen and Reynolds, 2002).
- Atom pairs are defined by:
 - the length of the shortest bond path between two atoms,
 - while the terminal atoms in this path are described by:
 - their element type
 - their number of pi electrons
 - their number of non-hydrogen neighbors
- Example: C12N03_06

Example



- Atom sequences:
 - similar to atom pairs, but all atoms in bond path are described.
 - Example: C12C13C13C02C02N03
- Conversion of atom pairs/sequences to binary vectors of constant length is usually not performed, but would be possible.

Similarity Coefficients

① Euclidean

$$\sqrt{\frac{c + d}{a + b + c + d}} \quad (1)$$

② Tanimoto coefficient

$$\frac{c}{a + b + c} \quad (2)$$

③ Simpson coefficient

$$\frac{c}{\min((a + c), (b + c))} \quad (3)$$

④ Tversky index

$$\frac{c}{\alpha * a + \beta * b + c} \quad (4)$$

⑤ Many more similarity coefficients (Holliday et al., 2003)

Legend for variables:

a: count of features in CMP A but not in CMP B

b: count of features in CMP B but not in CMP A

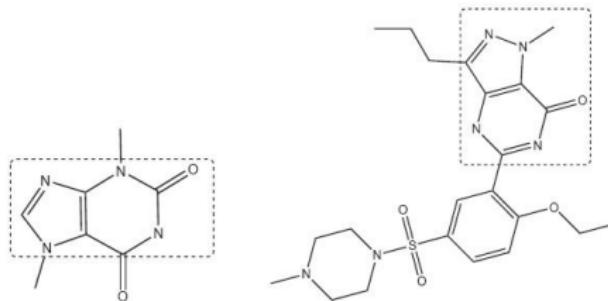
c: count of features in both CMP A and CMP B

d: count of features absent in CMP A and CMP B

α and β : weighting variables

MCS-based Similarity Concepts

- Graph-based algorithms that find maximum common substructure (MCS) shared among two molecules
- Flexible MCS matching algorithm implemented in *fmcsR* [Link](#) allows bond and atom mismatches.
- Major advantage: identification of local similarities



Alternatives: 3D Searches & Docking

Conformer Predictions

Prediction of the most stable conformers in 3D space.

3D Searches

Uses shape and topological indices to query a 3D conformer database.

3D Substructure searches

Related to pharmacophore searches

Docking

Computational modeling of the possible binding modes of a ligand to a target site.

Important Compound Databases

- PubChem
- DrugBank
- ChemBank
- ChEMBL
- Many more

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Compound Descriptors

Structural descriptors

- Atom pairs, fingerprints
- many others

Property descriptors

- Formula
- Molecular weight
- Octanol/Water partition coefficient ($\log P$)
- Hydrogen Bond Acceptors
- Hydrogen Bond Donors
- Acidic groups
- Rotatable bonds
- over 300-3000 additional ones

Drug-likeness Filters

Lipinski Rules

In a selection of 2245 compounds from the World Drug Index Lipinski identified four property cutoffs that were common in 90% of these drugs (Lipinski et al, 1997, *Adv Drug Deliv Rev*: 23, 3-25). These property filters are known as the "Rule of Five" (all multiple of 5):

- MW < 500g/mol
- lipophilicity: $\log P < 5$
- n H-bond donors < 5 (e.g. OH and NH)
- n H-bond acceptors < 10 (e.g. N and O)

Extended Lipinski Rules

- n rotatable bonds < 10

ADMET Rules

- Criteria for predicting adsorption, distribution, metabolism, excretion and toxicity (ADMET) more important for pharmaceutical industry than chemical genomics.

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Compound Structure Depictions

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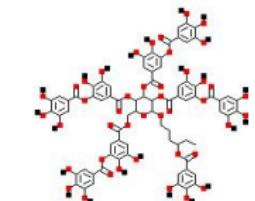
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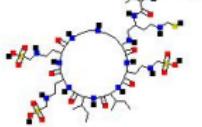
Clustering Methods

- Principal component analysis (PCA)
 - Reduction technique of multivariate data to principal components to identify hidden variances
- Multidimensional scaling
 - Displays distance matrix of objects in spacial plot
- Hierarchical Clustering
 - Iterative joining of items by decreasing similarity
- Jarvis-Patrick Clustering
 - Joins items based on intersects among nearest neighbor vectors
- Binning Clustering
 - Uses similarity cutoff for grouping of items
- Many additional clustering algorithms are being used in this field.

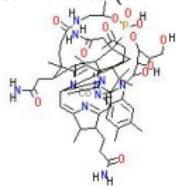
Example: PCA of Small Molecule Properties



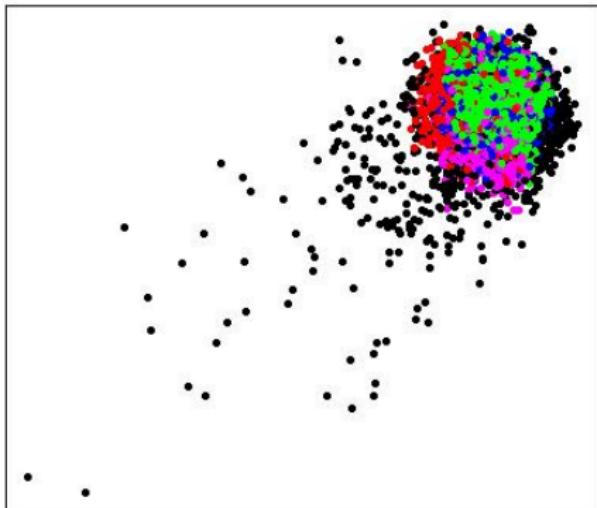
Tannic Acid (MW 1600)



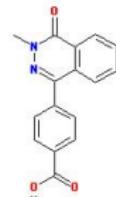
Colistimethate (MW 1400)



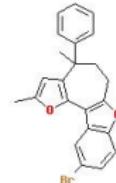
Cobalamin (MW 1100)



Comb1 Comb2 Comb3 Comb4 Bioact



56421 (MW 270)



19737 (MW 390)

Cheminformatics in R

Why cheminformatics in R?

- Open source
- Efficient data structures and graphics utilities
- Access to many clustering and machine learning algorithms
- Integration with bioscience packages

R packages for cheminformatics

- Bioconductor
 - *ChemmineR* [Link](#) (Cao et al., 2008)
 - *eiR* [Link](#)
 - *fmcsR* [Link](#)
 - *ChemmineOB*: R interface to subcomponents of *OpenBabel* [Link](#)
 - *ChemMine Tools* [Link](#): web interface to Chemmine utilities (Backman et al., 2011)
- CRAN
 - *rckd* [Link](#)
 - *rpubchem* [Link](#)

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Cheminformatics Basics

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Import of SD Files

Download R code for exercises and open in RStudio session

```
> download.file(url="http://faculty.ucr.edu/~tgirke/HTML_Presentations/Manuals/ChemmineR/Bioc2013/Cheminfo.R",
```

Loading the *ChemmineR* package and its documentation

```
> library("ChemmineR") # Loads the package
```

Accessing *ChemmineR* PDF manual and help documents

```
> library(help="ChemmineR") # Lists all functions and classes
```

```
> vignette("ChemmineR") # Opens PDF manual from R
```

```
> ?MW # Opens help for MW function
```

Import SD file into *SDFset* object

```
> sdfset <- read.SDFset("http://faculty.ucr.edu/~tgirke/Documents/R_BioCond/Samples/
```

```
> sdfset # Returns summary of SDFset
```

```
> valid <- validSDF(sdfset) # Identifies invalid SDFs in SDFset objects
```

```
> sdfset <- sdfset[valid] # Removes invalid SDFs, if there are any
```

Load sample SD file provided by package

```
> data(sdfsamp)
```

```
> sdfset <- sdfsamp
```

```
> sdfset # Returns summary of SDFset
```

An instance of "SDFset" with 100 molecules

Export Molecule Structures to SD Files

Write first 4 molecules to SD file

```
> write.SDF(sdfset[1:4], file="sub.sdf", sig=TRUE)
> list.files(pattern="sub.sdf")
[1] "sub.sdf"
```

Write all molecules to several files each containing 50 entries

```
> write.SDFsplit(x=sdfset, filetag="myfile", nmol=50)
      from   to      filename
1       1   50 myfile001_050.sdf
2     51  100 myfile051_100.sdf
```

Reimports newly created SD file

```
> sdfsetsub <- read.SDFset("sub.sdf")
> sdfsetsub
```

An instance of "SDFset" with 4 molecules

Exercise I: Import/Export

- Task 1 Open the PubChem site [Link](#) and search for 'p450 inhibitor'. Download the resulting 15 query hits to an SD file named 'p450.sdf' using the 'Structure Download' option on the right.
- Task 2 Import the 'p450.sdf' into your R session. Here is a backup [Link](#) of this file in case there are difficulties with the PubChem site.
- Task 3 Check in R the number of compounds stored in 'p450.sdf'.
- Task 4 Write the structures in inverted order back to an SD file.
- Task 5 Write the structures to several SD files each containing 5 molecules.

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Structure Formats

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Most Important S4 Objects in *ChemmineR*

Molecular structure containers

SDFstr: intermediate string class to facilitate SD file import; not important for end user

SDF: container for single molecule imported from an SD file

SDFset: container for many SDF objects; **most important container for end user**

Structure descriptor containers

AP: container for atom pair (AP) descriptors of a single molecule

APset: container for many AP objects

FP: container for fingerprint of a single molecule

FPset: container for fingerprints of many molecules

Important methods operating on *SDFset*-type containers

- Object slots: `cid`, `header`, `atomblock`, `bondblock`, `datablock`
- Structure depiction: `plot`

Coerce one class to another

- Standard syntax `as(..., "....")` works in most cases. For details see R help with `?"SDFset-class"`.

Working with SDF/SDFset Classes

Several methods are available to return the different data components of *SDF/SDFset* containers in batches. The following examples list the most important ones.

```
> view(sdfset[1:4]) # Summary view of several molecules  
> length(sdfset) # Returns number of molecules  
> sdfset[[1]] # Returns single molecule from SDFset as SDF object  
> sdfset[[1]][[2]] # Returns atom block from first compound as matrix  
> sdfset[[1]][[2]][1:4,]  
> c(sdfset[1:4], sdfset[5:8]) # Concatenation of several SDFsets
```

The `grepSDFset` function allows string matching/searching on the different data components of an *SDFset*. By default the function returns a SDF summary of the matching entries.

Alternatively, an index of the matches can be returned with the setting `mode="index"`.

```
> grepSDFset("650001", sdfset, field="datablock", mode="subset")  
>      # To return index, set mode="index")
```

Accessing SDF/SDFset Components

Methods for retrieving header, atom, bond and data blocks

```
> atomblock(sdf); sdf[[2]]; sdf[["atomblock"]]
>      # All three methods return the same component
> header(sdfset[1:4])
> atomblock(sdfset[1:4])
> bondblock(sdfset[1:4])
> datablock(sdfset[1:4])
```

Utilities to manage compound IDs and to keep them unique

```
> sdfid(sdfset[1:4])
[1] "650001" "650002" "650003" "650004"

>      # Retrieves CMP IDs from Molecule Name field in header block.
> cid(sdfset[1:4])
[1] "CMP1" "CMP2" "CMP3" "CMP4"

>      # Retrieves CMP IDs from ID slot in SDFset.
> unique_ids <- makeUnique(sdfid(sdfset))

[1] "No duplicates detected!"

>      # Creates unique IDs by appending a counter to duplicates.
> cid(sdfset) <- unique_ids # Assigns unqualified IDs to ID slot
```

Exercise II: SDFset Containers

- Task 1 Assign custom names to ID slot in `SDFset` and export object to SD file so that the custom IDs are used as IDs in the header block.
- Task 2 Extract the `bondblock` of all structures in an `SDFset`, `rbind` them with `do.call` and write the resulting `matrix` to a tabular file.
- Task 3 Replace `atomblock` of first molecule in `SDFset` with `atomblock` of second molecule. Check the result with "`==`".

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Compound Properties

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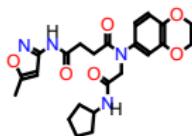
Compound Clustering

Rendering Chemical Structure Images

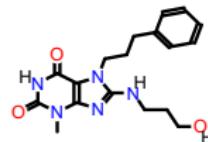
Plot compound Structures with R's graphics device

```
> data(sdfsamp); sdfset <- sdfsamp  
> plot(sdfset[1:4], print=FALSE) # print=TRUE returns SDF summaries
```

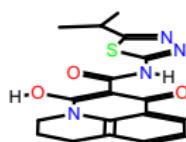
CMP1



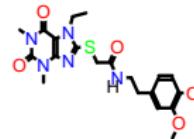
CMP2



CMP3



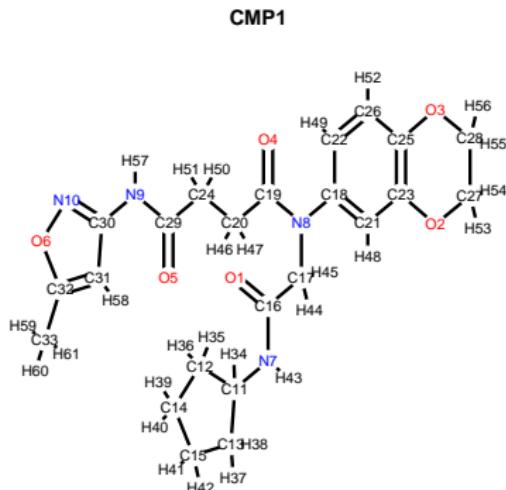
CMP4



Customize Structure Rendering

Show atom block position numbers next to the atom symbols. For more details, consult help documentation with `?plotStruc` or `?plot`.

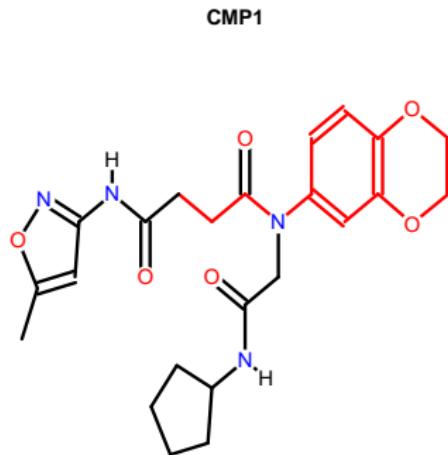
```
> plot(sdfset["CMP1"], atomnum = TRUE, noHbonds=F , no_print_atoms = "",  
+       atomcex=0.8, sub=paste("MW:", MW(sdfsmpale["CMP1"])), print=FALSE)
```



Substructure Coloring

Substructure highlighting by atom numbers

```
> plot(sdfset[1], print=FALSE, colbonds=c(22,26,25,3,28,27,2,23,21,18,8,19,20,24))
```



Online Structure Viewing with ChemMine Tools

Plot structures using web service ChemMine Tools:

```
> sdf.visualize(sdfset[1:4])
```

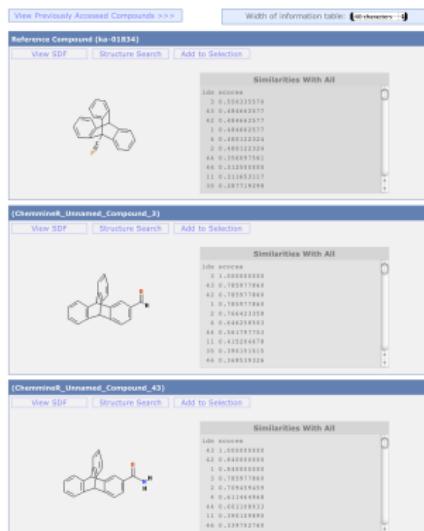


Figure: Visualization page created by calling `sdf.visualize`.

Exercise III: Rendering Compound Structures

- Task 1 Plot the structures of compound IDs "42631481", "42631375", "42631371" and "42631260" of the p450 SDFset that you created in Exercise I.
- Task 2 Render the same structures online with Chemmine Tools.

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Atom Count Table

Several methods and functions are available to compute basic compound descriptors, such as molecular formula (MF), molecular weight (MW), and frequencies of atoms and functional groups. In many of these functions, it is important to set addH=TRUE in order to include/add hydrogens that are often not specified in an SD file.

```
> propma <- atomcountMA(sdfset, addH=FALSE)
> propma[1:4,]
```

| | C | H | N | O | S | F | Cl |
|------|----|----|---|---|---|---|----|
| CMP1 | 23 | 28 | 4 | 6 | 0 | 0 | 0 |
| CMP2 | 18 | 23 | 5 | 3 | 0 | 0 | 0 |
| CMP3 | 18 | 18 | 4 | 3 | 1 | 0 | 0 |
| CMP4 | 21 | 27 | 5 | 5 | 1 | 0 | 0 |

Data frame provided by library containing atom names, atom symbols, standard atomic weights, group and period numbers.

```
> data(atomprop)
> atomprop[1:4,]
```

| Number | Name | Symbol | Atomic_weight | Group | Period |
|--------|-----------|--------|---------------|-------|--------|
| 1 | hydrogen | H | 1.007940 | 1 | 1 |
| 2 | helium | He | 4.002602 | 18 | 1 |
| 3 | lithium | Li | 6.941000 | 1 | 2 |
| 4 | beryllium | Be | 9.012182 | 2 | 2 |

Molecular Weight, Formula and Functional Groups

Compute MW and formula

```
> MW(sdfset[1:4], addH=FALSE)
```

| CMP1 | CMP2 | CMP3 | CMP4 |
|----------|----------|----------|----------|
| 456.4916 | 357.4069 | 370.4255 | 461.5346 |

```
> MF(sdfset[1:4], addH=FALSE)
```

| CMP1 | CMP2 | CMP3 | CMP4 |
|--------------|--------------|---------------|---------------|
| "C23H28N4O6" | "C18H23N5O3" | "C18H18N4O3S" | "C21H27N5O5S" |

Enumerate functional groups

```
> groups(sdfset[1:4], groups="fctgroup", type="countMA")
```

| | RNH2 | R2NH | R3N | ROPO3 | ROH | RCHO | RCOR | RCOOH | RCOOR | ROR | RCCH | RCN |
|------|------|------|-----|-------|-----|------|------|-------|-------|-----|------|-----|
| CMP1 | 0 | 2 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 0 | 0 |
| CMP2 | 0 | 2 | 2 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| CMP3 | 0 | 1 | 1 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0 | 0 |
| CMP4 | 0 | 1 | 3 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 0 | 0 |

Aggregate Many Molecular Properties

Combine MW, MF, charges, atom counts, functional group counts and ring counts in one data frame

```
> propma <- data.frame(MF=MF(sdfset, addH=FALSE), MW=MW(sdfset, addH=FALSE),
+                         Ncharges=sapply(bonds(sdfset, type="charge"), length),
+                         atomcountMA(sdfset, addH=FALSE), groups(sdfset,
+                                         type="countMA"), rings(sdfset, upper=6, type="count",
+                                         arom=TRUE))
> propma[1:4,]
```

| | MF | MW | Ncharges | C | H | N | O | S | F | Cl | RNH2 | R2NH | R3N | ROPO3 | ROH |
|------|-------------|----------|----------|-------|-----|------|-----|-------|---|----|------|------|-----|----------|-----|
| CMP1 | C23H28N4O6 | 456.4916 | | 0 | 23 | 28 | 4 | 6 | 0 | 0 | 0 | 2 | 1 | 0 | 0 |
| CMP2 | C18H23N5O3 | 357.4069 | | 0 | 18 | 23 | 5 | 3 | 0 | 0 | 0 | 2 | 2 | 0 | 1 |
| CMP3 | C18H18N4O3S | 370.4255 | | 0 | 18 | 18 | 4 | 3 | 1 | 0 | 0 | 0 | 1 | 1 | 0 |
| CMP4 | C21H27N5O5S | 461.5346 | | 0 | 21 | 27 | 5 | 5 | 1 | 0 | 0 | 0 | 1 | 3 | 0 |
| | RCHO | RCOR | RCOOH | RCOOR | ROR | RCCH | RCN | RINGS | | | | | | AROMATIC | |
| CMP1 | 0 | 0 | 0 | 0 | 2 | 0 | 0 | 4 | | | | | | 2 | |
| CMP2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 3 | | | | | | 3 | |
| CMP3 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 4 | | | | | | 2 | |
| CMP4 | 0 | 0 | 0 | 0 | 2 | 0 | 0 | 3 | | | | | | 3 | |

Assign Molecular Properties to SDF Data Block

The following shows an example for assigning the values stored in a matrix (e.g. property descriptors) to the data block components in an *SDFset*. Each matrix row will be assigned to the corresponding slot position in the *SDFset*.

```
> datablock(sdfset) <- propma # Works with all SDF components  
> datablock(sdfset)[1]
```

| MF | MW | Ncharges | C | H | N |
|--------------|------------|----------|------|-------|----------|
| "C23H28N4O6" | "456.4916" | "0" | "23" | "28" | "4" |
| 0 | S | F | Cl | RNH2 | R2NH |
| "6" | "0" | "0" | "0" | "0" | "2" |
| R3N | ROPO3 | ROH | RCHO | RCOR | RCOOH |
| "1" | "0" | "0" | "0" | "0" | "0" |
| RCOOR | ROR | RCCH | RCN | RINGS | AROMATIC |
| "0" | "2" | "0" | "0" | "4" | "2" |

Convert data block of *SDFset* to matrix.

```
> blockmatrix <- datablock2ma(datablocklist=datablock(sdfset))  
> blockmatrix[1:2,1:12]
```

| MF | MW | Ncharges | C | H | N | O | S | F | Cl | RNH2 | R2NH |
|-------------------|------------|----------|------|------|-----|-----|-----|-----|-----|------|------|
| CMP1 "C23H28N4O6" | "456.4916" | "0" | "23" | "28" | "4" | "6" | "0" | "0" | "0" | "0" | "2" |
| CMP2 "C18H23N5O3" | "357.4069" | "0" | "18" | "23" | "5" | "3" | "0" | "0" | "0" | "0" | "2" |

Charges and Missing Hydrogens

The function `bonds` returns information about the number of bonds, charges and missing hydrogens in *SDF* and *SDFset* objects. It is used by many other functions (e.g. `MW`, `MF`, `atomcount`, `atomcuntMA` and `plot`) to correct for missing hydrogens that are often not specified in SD files.

```
> bonds(sdfset[[1]], type="bonds")[1:4,]
```

| | atom | Nbondcount | Nbondrule | charge |
|---|------|------------|-----------|--------|
| 1 | 0 | 2 | 2 | 0 |
| 2 | 0 | 2 | 2 | 0 |
| 3 | 0 | 2 | 2 | 0 |
| 4 | 0 | 2 | 2 | 0 |

```
> bonds(sdfset[1:2], type="charge")
```

```
$CMP1  
NULL
```

```
$CMP2  
NULL
```

```
> bonds(sdfset[1:2], type="addNH")
```

```
CMP1 CMP2  
0 0
```

Ring Perception and Aromaticity Assignment

The function `rings` identifies all possible rings in one or many molecules using the exhaustive ring perception algorithm from Hanser et al. (1996). In addition, the function can return all smallest possible rings as well as aromaticity information.

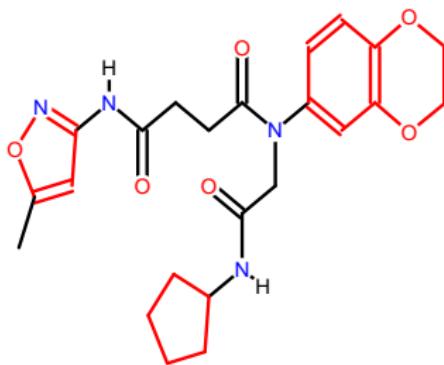
```
> (ringatoms <- rings(sdfset[1], upper=Inf, type="all", arom=TRUE, inner=FALSE))  
  
$RINGS  
$RINGS$ring1  
[1] "N_10" "O_6"  "C_32" "C_31" "C_30"  
  
$RINGS$ring2  
[1] "C_12" "C_14" "C_15" "C_13" "C_11"  
  
$RINGS$ring3  
[1] "C_23" "O_2"   "C_27" "C_28" "O_3"   "C_25"  
  
$RINGS$ring4  
[1] "C_23" "C_21" "C_18" "C_22" "C_26" "C_25"  
  
$RINGS$ring5  
[1] "O_3"   "C_28" "C_27" "O_2"   "C_23" "C_21" "C_18" "C_22" "C_26" "C_25"  
  
$AROMATIC  
ring1 ring2 ring3 ring4 ring5  
TRUE FALSE FALSE TRUE FALSE
```

Highlight Rings in Structure Image

For visual inspection, the corresponding compound structure can be plotted with the ring bonds highlighted in color.

```
> atomindex <- as.numeric(gsub(".*_", "", unique(unlist(ringatoms))))  
> plot(sdfset[1], print=FALSE, colbonds=atomindex)
```

CMP1



Streaming Through Large SD Files

The `sdfStream` function allows to stream through SD Files with millions of molecules without consuming much memory. During this process any set of descriptors, supported by *ChemmineR*, can be computed. In addition to descriptor values, the function returns a line index that gives the start and end positions of each molecule in the source SD File. This line index can be used by the downstream `read.SDFindex` function to retrieve specific molecules of interest from the source SD File without reading the entire file into R.

```
> write.SDF(sdfset, "test.sdf")
> desc <- function(sdfset) {
+   cbind(SDFID=sdfid(sdfset),
+         MW=MW(sdfset),
+         groups(sdfset),
+         rings(sdfset, type="count", upper=6, arom=TRUE)
+   )
+ }
> sdfStream(input="test.sdf", output="matrix.xls", fct=desc, Nlines=1000, silent=TRUE)
> read.delim("matrix.xls", row.names=1)[1:3,1:10]
```

| | SDFlineStart | SDFlineEnd | SDFID | MW | RNH2 | R2NH | R3N | ROP03 | ROH | RCHO |
|------|--------------|------------|-------|--------|----------|------|-----|-------|-----|------|
| CMP1 | | 1 | 203 | 650001 | 456.4916 | 0 | 2 | 1 | 0 | 0 |
| CMP2 | | 204 | 381 | 650002 | 357.4069 | 0 | 2 | 2 | 0 | 1 |
| CMP3 | | 382 | 550 | 650003 | 370.4255 | 0 | 1 | 1 | 0 | 1 |

Exercise IV: Compound Properties

- Task 1 Compute for p450 SDFset from Exercise I all possible compound properties.
- Task 2 Assign the property matrix to the data block in the corresponding SDFset.
- Task 3 Export the modified SDFset object to an SD file and inspect the result.

Outline

Cheminformatics Basics

Structure Formats

Similarity Searching

Physicochemical Properties

Clustering

Hands-on Section

Compound Import/Export

Object Classes

Compound Structure Depictions

Compound Properties

Compound Similarity Searching

Compound Clustering

Structure Descriptor Containers: APset/FPset

The function `sdf2ap` computes atom pair descriptors for one or many compounds (Chen and Reynolds, 2002; Cao et al., 2008). It returns a searchable atom pair database stored in a container of class `APset`, which can be used for structural similarity searching and clustering.

```
> apset <- sdf2ap(sdfset)
> apset
```

An instance of "APset" with 100 molecules

Most methods working on SDFset objects work the same way on descriptor objects.

```
> showClass("APset")
> cid(apset)
> view(apset)
> as(apset, "list")
```

The `FPset` class stores fingerprints of small molecules in a matrix-like representation where every molecule is encoded as a fingerprint of the same type and length.

```
> (fpset <- desc2fp(apset))
```

An instance of a 1024 bit "FPset" with 100 molecules

```
> view(fpset[1])
```

\$CMP1

An instance of "FP"

```
<<fingerprint>>
```

```
0 0 0 0 0 1 0 1 1 1 0 1 0 0 1 0 0 0 0 0 ... length: 1024
```

Atom Pair and Atom Pair Fingerprint Searches

The `cmp.search` function searches an atom pair database for compounds that are similar to a query compound.

```
> cmp.search(apset, apset["CMP1"], type=3, cutoff = 0.3, quiet=TRUE)
```

| | index | cid | scores |
|---|-------|-------|-----------|
| 1 | 1 | CMP1 | 1.0000000 |
| 2 | 96 | CMP96 | 0.3516643 |
| 3 | 67 | CMP67 | 0.3117569 |
| 4 | 88 | CMP88 | 0.3094629 |
| 5 | 15 | CMP15 | 0.3010753 |

Compound similarity searching with *FPset*

```
> fpset1024 <- names(rev(sort(table(unlist(as(apset, "list")))))[1:1024])
```

```
> fpset <- desc2fp(apset, descnames=fpset1024, type="FPset")
```

```
> fpSim(fpset["CMP1"], fpset, method="Tanimoto", cutoff=0.2, top=6)
```

| CMP1 | CMP96 | CMP67 | CMP31 | CMP88 | CMP15 |
|-----------|-----------|-----------|-----------|-----------|-----------|
| 1.0000000 | 0.4300000 | 0.3859060 | 0.3855856 | 0.3804714 | 0.3738602 |

Similarity Searching with PubChem Fingerprints

The `fpSim` function can be used for any type of binary fingerprint, here PubChem fingerprints extracted from the data block of the `SDFset` object.

```
> fpset <- fp2bit(sdfsample, type=3)
> fpSim(fpset[1], fpset, method="Tanimoto", cutoff=0.2, top=6)
```

| CMP1 | CMP33 | CMP98 | CMP86 | CMP4 | CMP70 |
|-----------|-----------|-----------|-----------|-----------|-----------|
| 1.0000000 | 0.6950000 | 0.6763485 | 0.6666667 | 0.6448980 | 0.6422018 |

Pairwise comparisons are supported as well

```
> fpSim(fpset[1], fpset[2])
```

| CMP2 |
|-----------|
| 0.5364807 |

Maximum Common Substructure (MCS) Searching

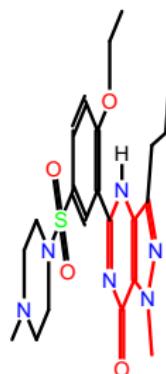
The *fmcsR* package provides support for identifying strict MCSs and mismatch tolerant flexible MCSs among compounds.

```
> library(fmcsR); data(fmcstest) # Loads library and test sdfset object  
> test <- fmcs(fmcstest[1], fmcstest[2], au=2, bu=1) # Searches for MCS with mismatch  
> plotMCS(test) # Plots both query compounds with MCS in color
```

Caffeine



Viagra



FMCS-based structure similarity searching

```
> fmcsBatch(sdfset[[1]], sdfset)[1:2,]
```

| | Query_Size | Target_Size | MCS_Size | Tanimoto_Coefficient | Overlap_Coefficient |
|------|------------|-------------|----------|----------------------|---------------------|
| CMP1 | 33 | 33 | 33 | 1.0000000 | 1.0000000 |
| CMP2 | 33 | 26 | 11 | 0.2291667 | 0.4230769 |

Searching PubChem from ChemmineR

ChemmineR supports searching of the PubChem database by compound IDs or via a structure similarity search using PubChem fingerprints. The following searches PubChem by structure similarity and stores the results in an SDFset object.

```
> compounds <- searchSim(sdfset[1])  
> compounds
```

An instance of "SDFset" with 10 molecules

Exercise V: Compound Similarity Searching

- Task 1** Convert the p450 SDFset from Exercise I into 3 searchable descriptor databases containing: (1) atom pairs, (2) atom pair fingerprints and (3) PubChem fingerprints.
- Task 2** Perform a structure similarity search against all three databases with the first compound in p450 SDFset as query. Compare the ranking of the three different search results.

Outline

Cheminformatics Basics

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Binning Clustering

Compound libraries can be clustered into discrete similarity groups with the binning clustering function `cmp.cluster`.

```
> c1 <- cmp.cluster(fpset, cutoff=c(0.3, 0.6, 0.9), method="Tanimoto",
+                      quiet=TRUE)[1:8,]
```

sorting result...

```
> c1[1:8,]
```

| | ids | CLSZ_0.3 | CLID_0.3 | CLSZ_0.6 | CLID_0.6 | CLSZ_0.9 | CLID_0.9 |
|---|------|----------|----------|----------|----------|----------|----------|
| 1 | CMP1 | 100 | 1 | 91 | 1 | 1 | 1 |
| 2 | CMP2 | 100 | 1 | 91 | 1 | 1 | 2 |
| 3 | CMP3 | 100 | 1 | 91 | 1 | 1 | 3 |
| 4 | CMP4 | 100 | 1 | 91 | 1 | 1 | 4 |
| 6 | CMP6 | 100 | 1 | 91 | 1 | 1 | 6 |
| 7 | CMP7 | 100 | 1 | 91 | 1 | 1 | 7 |
| 8 | CMP8 | 100 | 1 | 91 | 1 | 1 | 8 |
| 9 | CMP9 | 100 | 1 | 91 | 1 | 1 | 9 |

```
> cluster.sizestat(c1, cluster.result=2)
```

| | cluster | size | count |
|---|---------|------|------------|
| 1 | | 91 | 0.08791209 |

Jarvis-Patrick Clustering

The Jarvis-Patrick clustering algorithm is widely used in cheminformatics (Jarvis and Patrick, 1973) because it scales to very large numbers of compounds. The following performs standard Jarvis-Patrick clustering and computes the nearest neighbor table on the fly.

```
> jarvisPatrick(nearestNeighbors(fpset, numNbrs=6), k=5, mode="a1a2b") [1:20]
```

| CMP1 | CMP2 | CMP3 | CMP4 | CMP5 | CMP6 | CMP7 | CMP8 | CMP9 | CMP10 | CMP11 | CMP12 | CMP13 |
|-------|-------|-------|-------|-------|-------|-------|------|------|-------|-------|-------|-------|
| 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 |
| CMP14 | CMP15 | CMP16 | CMP17 | CMP18 | CMP19 | CMP20 | | | | | | |
| 14 | 15 | 16 | 17 | 18 | 19 | 20 | | | | | | |

Output nearest neighbor table (*matrix*)

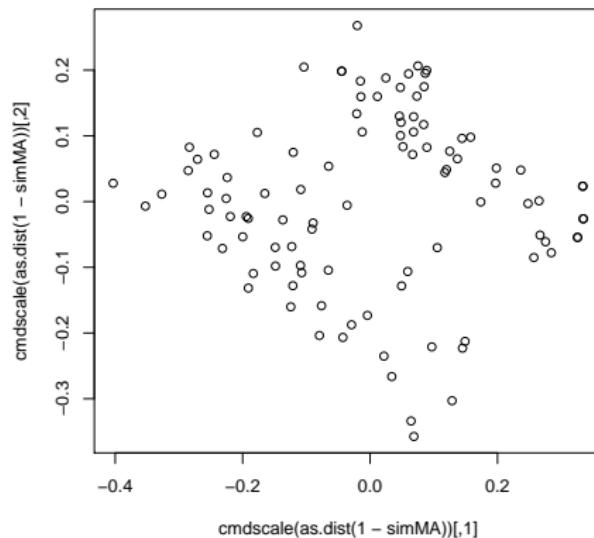
```
> nnm <- nearestNeighbors(fpset, numNbrs=6)
> nnm$similarities[1:4,]
```

| | CMP1 | CMP33 | CMP98 | CMP86 | CMP4 | CMP70 |
|-----|------|-----------|-----------|-----------|-----------|-----------|
| sim | 1 | 0.6950000 | 0.6763485 | 0.6666667 | 0.6448980 | 0.6422018 |
| sim | 1 | 0.7823834 | 0.7475248 | 0.7348837 | 0.7281553 | 0.6666667 |
| sim | 1 | 0.6871795 | 0.6446701 | 0.6283186 | 0.6276596 | 0.6263158 |
| sim | 1 | 0.8177570 | 0.7348837 | 0.7287449 | 0.7136564 | 0.7105263 |

Multi-Dimensional Scaling (MDS)

Multidimensional scaling (MDS) algorithms start with a matrix of item-item distances and then assign coordinates for each item in a low-dimensional space to represent the distances graphically.

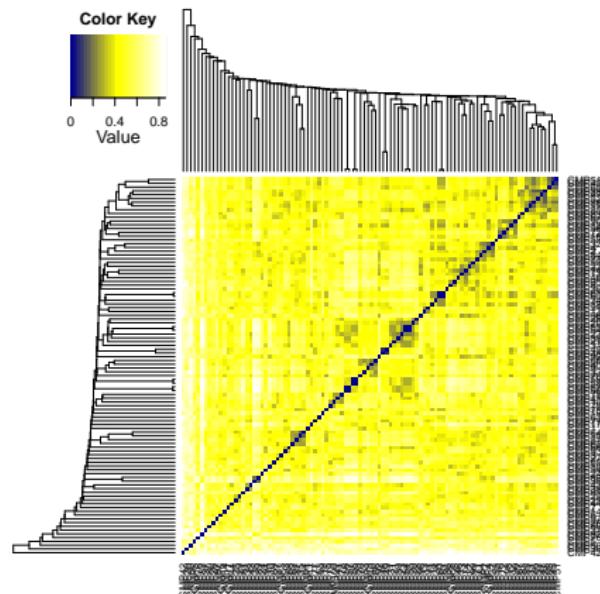
```
> simMA <- sapply(cid(fpset), function(x) fpSim(fpset[x], fpset, sorted=FALSE))  
> plot(cmdscale(as.dist(1-simMA)))
```



Hierarchical Clustering

The following performs hierarchical clustering of compound structure similarities (distances). The resulting dendrogram is then plotted next to a heatmap of the corresponding similarity matrix.

```
> library(gplots)
> hc <- hclust(as.dist(1-simMA), method="single")
> heatmap.2(1-simMA, Rowv=as.dendrogram(hc), Colv=as.dendrogram(hc),
+           col=colorpanel(40, "darkblue", "yellow", "white"),
+           density.info="none", trace="none")
```



Exercise VI: Compound Clustering

- Task 1 Cluster the structures in p450 SDFset with the binning clustering algorithm.
- Task 2 Cluster the structures in p450 SDFset with the Jarvis-Patrick clustering algorithm.
- Task 3 Cluster the structures in p450 SDFset with the MDS algorithm.
- Task 4 Cluster the structures in p450 SDFset with the hierarchical clustering algorithm.

Session Information

```
> sessionInfo()

R version 3.0.0 (2013-04-03)
Platform: x86_64-pc-linux-gnu (64-bit)

locale:
[1] LC_CTYPE=en_US.UTF-8      LC_NUMERIC=C
[3] LC_TIME=en_US.UTF-8       LC_COLLATE=en_US.UTF-8
[5] LC_MONETARY=en_US.UTF-8   LC_MESSAGES=en_US.UTF-8
[7] LC_PAPER=C                LC_NAME=C
[9] LC_ADDRESS=C              LC_TELEPHONE=C
[11] LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C

attached base packages:
[1] grid      stats     graphics grDevices utils     datasets methods
[8] base

other attached packages:
[1] gplots_2.11.0.1    MASS_7.3-26        KernSmooth_2.23-10 caTools_1.14
[5] gdata_2.12.0.2     gtools_2.7.1       fmcsR_1.2.0        ChemmineR_2.12.1

loaded via a namespace (and not attached):
[1] bitops_1.0-5      DBI_0.2-5        digest_0.6.3      RCurl_1.95-4.1 tools_3.0.0
```

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